

1 **Quantifying the impact of nevirapine-based prophylaxis strategies to**
2 **prevent mother-to-child transmission of HIV-1:**
3 **A combined pharmacokinetic, pharmaco- and viral dynamic analysis**
4 **to predict clinical outcomes**

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15 **Running title:** Impact of NVP prophylaxis on vertical transmission of HIV-1

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24 women and newborns, virus dynamics, HIV transmission risk, drug resistance,
25 mathematical modeling

26 **Abstract**

27 Nevirapine single dose (sd-NVP) and extended NVP prophylaxis are widely used in
28 resource-constrained settings to prevent vertical HIV-1 transmission.

29 We assessed the pharmacokinetics of sd-NVP in 62 HIV-1 positive Ugandan pregnant
30 woman and their newborns, taking sd-NVP prophylaxis to prevent mother-to-child HIV-1
31 transmission. Based on this data we developed a mathematical model system to
32 quantify the impact of different sd-NVP regimens at delivery and extended infant NVP
33 prophylaxis (6, 14, 21, 26, 52, 78, 102 weeks) on the 2 year risk of HIV-1 transmission
34 and development of drug resistance in mothers and their breast-fed infants.

35 Pharmacokinetic parameter estimates and model-predicted HIV-1 transmission rates
36 were very consistent with other studies. Predicted 2 year HIV-1 transmission risks were
37 35.8% without prophylaxis, 31.6% for newborn sd-NVP, 19.1% for maternal sd-NVP and
38 19.7% for maternal/newborn sd-NVP. Maternal sd-NVP reduced newborn infection
39 predominately by trans-placental exchange, providing protective NVP concentrations to
40 the newborn at delivery, rather than by maternal viral load reduction. Drug resistance
41 was frequently selected in HIV-1 positive mothers after maternal sd-NVP.

42 Extended newborn NVP prophylaxis further decreased HIV-1 transmission risks but
43 indicated an overall decline in cost-effectiveness for increasing durations of newborn
44 prophylaxis. The total number of newborn infections with resistant virus was not
45 increased by extended newborn NVP prophylaxis.

46 The developed mathematical modeling framework successfully predicted the risk of HIV-
47 1 transmission and resistance development and can be adapted to other drugs/drug
48 combinations to *a priori* assess their potential in reducing vertical HIV-1 transmission
49 and resistance spread.

50 **Introduction**

51 HIV-1 infection remains a serious health care problem worldwide. In 2009,
52 approximately 370,000 children became infected with HIV-1 (53). Mother-to-child
53 transmission rates of HIV-1 in untreated breastfeeding populations in resource-limited
54 settings ranged from 25% to 48%, accounting for the vast majority of pediatric AIDS (12).

55 Vertical transmission of HIV-1 may occur during pregnancy (5%-10%), during birth
56 (10%-20%), and via breastfeeding (10%-20%) (12).

57 Intrapartum and newborn single dose nevirapine (NVP) significantly reduce transmission
58 of HIV-1 from the mother to the child (23) and are essential components of HIV-1
59 prevention strategies in many resource-constrained settings (57-58). However, the exact
60 mechanism of HIV-1 prevention by NVP during intrapartum transmission remains
61 unknown. Furthermore, owing to its long half-life, NVP frequently selects drug resistant
62 viral strains in HIV-infected mothers (17, 22), which can compromise the efficacy of
63 follow-up maternal and newborn antiretroviral treatment (ART) (8, 25, 29, 39).

64 In many resource-constrained settings, breastfeeding is critical for infant survival (59).
65 Reduction of HIV-1 transmission by short-course antiviral prophylaxis is frequently
66 impaired by subsequent infection during the breastfeeding period (24, 42). Extended
67 newborn NVP prophylaxis has shown to reduce HIV-1 transmission via breastfeeding (3,
68 6, 27, 38) and current WHO guidelines for the prevention of mother-to-child transmission
69 recommend the use of NVP throughout the entire breastfeeding period (57), which can
70 be as long as 2 years. Clinical trial data on extended prophylactic newborn NVP are
71 currently only available for durations of 6 weeks and 6 months (3, 10, 38). However, to
72 evaluate the effectiveness of extended newborn NVP, a quantification of the HIV-1

73 transmission risks after different durations of extended NVP prophylaxis in newborns is
74 required.

75 In the present study, NVP plasma data of 62 Ugandan mothers and newborns who took
76 NVP single dose prophylaxis were simultaneously analyzed in a single integrated
77 population pharmacokinetic model for both populations; the present work extends a
78 previously published pharmacokinetic study, which analyzed woman and newborn NVP
79 concentrations separately (28). The aim of this work was to combine pharmacokinetic
80 and pharmacodynamic analysis by developing a single mathematical modeling
81 framework. The framework should be used to predict the impact of various single and
82 extended NVP-based prophylaxis regimens on the cumulative risk of vertical HIV-1
83 transmission and on selection of NVP-resistant virus.

84 **Methods**

85 *Patient Characteristics and Study Design*

86 During a program for the prevention of mother-to-child transmission of HIV-1 in western
87 Uganda, 62 HIV-1 positive pregnant women and their newborns were enrolled for
88 pharmacokinetic (PK) analysis after they had given informed consent and delivered at
89 Fort Portal District Hospital (Fort Portal, Kabarole District, western Uganda). Pregnant
90 women received a single 200 mg NVP tablet at onset of labor and newborns received
91 2 mg/kg NVP syrup orally within 72 h after birth (28). Ethical approval was obtained from
92 the Uganda National Council for Science and Technology.

93 Median age and body weight of the pregnant women were 26 years and 56 kg [range:
94 16-39 yrs; 42-84 kg], respectively. Newborns had a median body weight of 3.1 kg [range:
95 2.0-3.9 kg]. The median time period passed between NVP intake of pregnant women

96 and birth was 5.1 h [range: 0.3-24.8 h]. The median time interval between birth and NVP
97 administration to the newborn was 0.9 h [range: 0.1-40.6 h] and 8.5 h [range: 1.3-46 h]
98 from NVP intake of the pregnant women until the NVP administration to the newborns
99 (28).

100 For PK analysis, a total of 113 plasma samples from mothers and newborns were
101 collected over three time periods, i.e. delivery, week 1 and week 2. The geometric mean
102 NVP concentration-time profile was previously presented (28). Here we illustrate the
103 dispersion of the individual plasma concentrations over time for the same population
104 (Figure 1 A). NVP concentrations were determined by a validated LC/tandem-mass-
105 spectrometry method according to the criteria set by the FDA (FDA-Guideline, (28, 48)).

106 *Pharmacokinetic Analysis*

107 Based on the previously established pharmacokinetic models and data (28), an
108 integrated population pharmacokinetic model was developed to simultaneously analyze
109 NVP plasma data of mothers and newborns.

110 For population PK data analysis the nonlinear mixed effects modeling approach
111 implemented in the software program NONMEM™ (Icon development solutions, version
112 VI, 1st update, 2006) was chosen due to sparse data situation. The pharmacokinetic
113 model was parameterized in terms of clearance(s) (CL) and volumes(s) of distribution (V)
114 using the PREDPP subroutines (FOCE with interaction, ADVAN6 TOL5) supplied in
115 NONMEM™. The model building process was guided by changes in the objective
116 function value of nested models provided by NONMEM™, by precision of the PK
117 parameter estimates (relative standard errors, RSE) and by basic goodness of fit (GOF)
118 plots. In addition, 2000 bootstrap data sets were assessed using NONMEM™. For

119 model evaluation, the final PK parameter estimates were compared to the corresponding
120 median and 95% confidence interval of the bootstrap runs. Model-based simulations for
121 visual predictive checks (VPC) were performed by NONMEM™ (n=1000 simulations)
122 and the statistics of 5th, median and 95th percentile were calculated using R, version 2.9.
123 The schematic structure of the final PK model for maternal and newborn data is
124 presented in Figure 1 B. Due to the difference in drug transport processes, solid lines
125 represent those occurring continuously over the whole time and dashed lines only those
126 before delivery except K34 which occurs only after delivery. The NVP absorption rate
127 constants for pregnant women (KA) and newborns (K34) were fixed to 1.34 h⁻¹,
128 respectively, due to no available data during the absorption process. Prior published
129 values of absorption rates varied between 0.013 h⁻¹ and 3.81 h⁻¹ (median: 1.3 h⁻¹) (4, 11,
130 16, 26).

131 Maternal plasma concentrations were associated to the central compartment with the
132 volume of distribution V2 and fetal/newborn concentrations to the peripheral
133 compartment with the volume of distribution V4. After delivery, but before the NVP
134 administration to newborns, significant NVP concentrations were detected in the plasma
135 of newborns. Considering the placenta-permeability of NVP (35, 37), we implemented a
136 trans-placental exchange of NVP between pregnant woman and fetus (PCL) before the
137 time of delivery (dashed lines in Figure 1 B). The ratio of NVP plasma concentrations
138 between fetus and pregnant women was described by a partition coefficient, PCM. NVP
139 elimination from the central compartment (related to the plasma of mothers) and the
140 peripheral compartment (related to the plasma concentration in the fetus/newborn) were
141 described by the PK parameters CL1 and CL2, respectively.

142 For the predictive performance of the final PK model a visual predictive check (VPC) is
 143 depicted in Figure 2 A and B. The dashed lines represent the 5th and 95th model-
 144 simulated percentiles and solid lines represent the model-simulated median of NVP
 145 concentrations. The VPC of mother plasma (Figure 2 A) and newborn plasma data
 146 (Figure 2 B) revealed sufficient model predictive performance for the general trend.
 147 Overall, the model-predicted variability was sufficient for mothers and newborns and
 148 resembled the variability in the observed data.

149 *HIV-1 Dynamics Model*

150 In order to quantify the impact of NVP prophylaxis on virus transmission, we adapted the
 151 virus dynamics model presented in (55) by discarding the longer lived cell types
 152 (representing macrophages and latently infected T-cells), as they do not impact the
 153 observed viral dynamics after short course maternal NVP. The utilized model of HIV-1
 154 dynamics and mother-to-child transmission is depicted in Figure 3 A.

155 Briefly, the mathematical model of virus dynamics and mutation comprises T-cells T ,
 156 free virus V , early infected T-cells T₁ (after reverse transcription but before viral
 157 genomic integration) and productively infected T-cells T₂ (after viral genomic integration).

158 The average rate of change of the different T-cell species and the number of viruses is
 159 given by the following system of ordinary differential equations:

$$160 \quad \frac{d}{dt} T = \lambda(t) - T \cdot \delta_T - \sum_i \beta(i, t) \cdot V(i) \cdot T + \sum_i \delta_{PIC} \cdot T_1(i) \quad (1)$$

$$161 \quad \frac{d}{dt} T_1(i) = \sum_j p_{j \rightarrow i} \cdot \beta(j, t) \cdot V(j) \cdot T - T_1(i) \cdot (\delta_{T_1} + k_T + \delta_{PIC}) \quad (2)$$

$$162 \quad \frac{d}{dt} T_2(i) = k_T \cdot T_1(i) - T_2(i) \cdot \delta_{T_2} \quad (3)$$

163
$$\frac{d}{dt} V(i) = N \cdot T_2(i) - V \cdot CL_V(t) \quad (4)$$

164 In summary, free virus V of strain i can infect T-cells with infection rate constant β ,
 165 which encompasses all steps from target cell binding, fusion, to reverse transcription,
 166 resulting in early infected cells T_1 , which turn into productively infected cells T_2 by
 167 provirus translocation into the nucleus and integration with rate k_T . T_2 produce new
 168 virus V with the rate constant N (on average 1000 virions/day/cell (45)). Native, early
 169 infected and productively infected T-cells are degraded with rate constants δ_T, δ_{T_1} and
 170 δ_{T_2} , respectively. In early infected cells T_1 (prior to proviral integration) essential
 171 components of the pre-integration complex can be degraded with rate constant δ_{PIC} ,
 172 returning the cell to an uninfected stage T (55). Native T-cells are produced with rate
 173 constant $\lambda(t)$ and free virus V is cleared with rate $CL_V(t)$ by the immune system. We
 174 assumed that the rate constants $\lambda(t)$ and $CL_V(t)$ are constant for the HIV-infected
 175 mothers, whereas they were considered time-dependent for the newborn, due to
 176 immune system development and growth. A derivation of the parameters $\lambda(t)$ (newborn)
 177 and $CL_V(t)$ (newborn) is provided in the supplementary text S1. All model parameters
 178 are displayed in Table 1.

179 **Viral Mutation:**

180 HIV can acquire drug resistance by mutation during the process of reverse transcription
 181 (comprised in parameter β in the model). The probability that a specific mutation occurs
 182 during the process of reverse transcription has been quantified *ex vivo* to be
 183 $\mu = 2.16 \cdot 10^{-5}$ (per base and reverse transcription process) (30). A single genomic point

184 mutation inducing a change at the protein level, e.g. position Y181 -> 181C (Y181C) will
 185 therefore occur with probability μ during reverse transcription, whereas with probability
 186 $(1-\mu)$ this specific mutation will not occur. In our model, 2 specific sites L are regarded
 187 to undergo mutation; resulting in the K103N and the Y181C change in the reverse
 188 transcriptase enzyme, respectively. As an example, the probability that the wild type
 189 virus wt will neither be mutated at one of the 2 sites is $p_{wt \rightarrow wt} = (1-\mu)^2$. The probabilities
 190 that precisely one mutation occurs is given by $p_{wt \rightarrow Y181C} = (1-\mu) \cdot \mu$ and the probability of
 191 two specific mutations by $p_{wt \rightarrow K103N/Y181C} = \mu^2$. More generally, the probability that a
 192 certain transition by mutation from some strain j to some strain i occurs during reverse
 193 transcription $p_{j \rightarrow i}$, is given by:

$$194 \quad p_{j \rightarrow i} = \mu^{h(i,j)} \cdot (1-\mu)^{L-h(i,j)}, \quad (5)$$

195 where $h(i, j)$ denotes the hamming distance (the number of differences) between strain
 196 j and strain i . All mutation probabilities for the utilized model are depicted in Figure 3 B.

197 Coupling of Viral Dynamics with NVP Pharmacokinetics:

198 The efficacy of the non-nucleoside reverse transcriptase inhibitor NVP ($1-\eta(i, t)$) at time
 199 t against strain i was implemented using the standard E_{\max} -model with slope parameter
 200 (44, 47):

$$201 \quad (1-\eta(i, t)) = \frac{1}{1 + \left(\frac{C(t)}{IC_{50}(i)} \right)^{h(i)}} \quad (6)$$

202 where $C(t)$ denotes the NVP concentration at time t (derived during PK analysis, see
 203 above), $IC_{50}(i)$ denotes the strain-specific fifty percent inhibitory concentration and $h(i)$

204 denotes the strain-specific slope parameter (see Figure 3 C). The strain specific
205 infection rate constant under treatment was given by
206 $\beta(i,t) = (1-\eta(i,t)) \cdot (1-s(i)) \cdot \beta(wt,\phi) \cdot SF(t)$, where $\beta(wt,\phi)$ denotes the infection rate
207 constant of the wild type wt in the absence of drug ϕ (given in Table 1) and $s(i)$,
208 denotes the fitness loss (e.g., loss in the activity of reverse transcriptase) relative to the
209 wild type (shown in Figure 3 C). The scaling factor $SF(t)$ corrects the infection rate β for
210 the differences in target cell concentration between mother (reference target cell
211 concentration) and uninfected newborn. $SF(t)$ was considered to be time-dependent for
212 newborns due to immune system development and growth (see Equation S3 and Figure
213 S1 of the supplementary material), whereas it was set to the value of 1 for HIV-1
214 infected mothers.

215 *Deterministic –Stochastic Hybrid Simulation*

216 The kinetics of biological systems, in which all reactions occur quasi-continuously over
217 time or involve large numbers of reactants are well approximated by continuous-
218 deterministic simulations (by numerical solution of the systems' ordinary differential
219 equations). However, the exact kinetics of biological systems which involve rare reaction
220 events with small numbers of reactants are intrinsically stochastic and are therefore only
221 poorly approximated by continuous-deterministic simulation (61). In our modelling
222 framework, the process of HIV-1 transmission denotes such an event in which the
223 outcome is intrinsically stochastic: Either the transmitted virus becomes entirely cleared
224 by the immune system before establishing stable infection ($V = 0$), or it succeeds in
225 establishing infection (V approaches its steady state).

226 In order to fully regard the intrinsic stochasticity of rare events in the utilized model (such
227 as viral challenges) and to allow efficient simulation of quasi-continuous kinetics, we
228 chose the deterministic-stochastic hybrid simulation approach presented in (1).

229 *Prediction of HIV-1 Transmission after Intrapartum- and Extended NVP* 230 *Prophylaxis*

231 Model simulations were performed starting with the time of the first maternal dose (if no
232 dose was given, with the time of birth) and continued until 2 years postpartum. The
233 number of viruses coming into contact with the newborn during delivery and
234 breastfeeding was modeled as a function of the maternal viral load at the particular time
235 of the respective viral challenge. The intrapartum virus transmission was modeled in
236 terms of a single viral challenge at the time of delivery, while virus transmission via
237 breastfeeding was modeled in terms of repeated viral challenges during the time after
238 delivery until 2 years postpartum. The probability of a viral challenge during
239 breastfeeding was assumed to decrease over time (see Figure S2, supplementary
240 material). Child growth and immune system development were considered
241 simultaneously to the model simulation (see supplementary text S1 and Figure S1,
242 supplementary material). If stable infection of the newborns occurred (defined as total
243 number of viruses $\geq 1 \cdot 10^6$ in the newborn) the respective simulation was stopped and
244 the time between birth and child infection was recorded for subsequent evaluation. The
245 cumulative infection risk 2 years postpartum was assessed by Kaplan-Meier estimates
246 and an intrauterine transmission probability of 5% (12) was added. All model predictions
247 are based on 1000 hybrid deterministic-stochastic simulations to ensure statistic
248 confidence in the results.

249 We considered four scenarios for sd-NVP: (A: no prophylaxis, B: single postpartum
250 newborn 2 mg/kg NVP dose, C: single intrapartum maternal 200 mg NVP dose and D:
251 intrapartum maternal 200 mg NVP dose plus postpartum newborn 2 mg/kg NVP dose).
252 We took into account the patient characteristics from the Ugandan program for the
253 prevention of mother-to-child transmission discussed above, in particular the individual
254 time intervals between maternal NVP administration and birth; median 5.1 h [range: 0.3-
255 24.8 h] and the time intervals between birth and newborn NVP administration; median
256 0.9 h [range: 0.1-40.6 h].

257 For the *extended* newborn NVP prophylaxis, we first simulated HIV-1 dynamics with
258 maternal intrapartum NVP plus one postpartum newborn NVP dose, as described above,
259 until day 1 after birth, after which we simulated HIV-1 dynamics until 2 years postpartum,
260 following either 6 weeks (SWEN-study (3, 38)), 14-, 21 weeks, 6 months (HPTN 046-
261 study (10)), 52-, 78- or 104 weeks of daily oral 2 mg/kg NVP administration, taking into
262 account the pharmacokinetic characteristics of the population in the program for the
263 prevention of mother-to-child transmission.

264 **Results**

265 *Pharmacokinetics of NVP in Pregnant Women/Mothers and Their* 266 *Newborns*

267 The estimated PK parameters (using the model in Figure 1 B) are presented in Table 2.
268 Mother and newborn data were best described by combined 1-compartment models with
269 first-order absorption and elimination processes. Since the bioavailability of the oral dose
270 was unknown, the estimated PK parameters have to be reported as relative parameters.
271 The relative volume of distribution of mother data, V_2/F was estimated to be 90.9 L and

272 the relative NVP clearance to be 1.22 L/h. Inter-individual variabilities (IIV) were
273 implemented for all structural parameters relating to mother data and estimated to be
274 moderate (34% and 33% coefficient of variation (CV) for V_2/F and CL_1/F , respectively)
275 but high for KA (160% CV). The placenta clearance PCL/F was estimated to be 111 L/h
276 suggesting a rapid placental transfer. The partition coefficient between NVP
277 concentrations of fetus and pregnant women (PCM) was quantified to be 1.38. The large
278 volume of distribution and low elimination capacity resulted in a long half-life of 52 h for
279 mothers. The relative volume of distribution V_4/F for newborns was estimated to be
280 20.0 L and the relative clearance CL_2/F to be 0.21 L/h. The half-life of NVP in newborns
281 was 66 h.

282 The residual variability was best described using separate proportional error models for
283 maternal and newborn data, respectively. The proportional error was moderate (27% CV)
284 for mother and higher (49% CV) for newborn data. The precision of the estimated PK
285 parameters was sufficient with $RSE < 20.5\%$ for fixed-effects and $RSE < 33\%$ for
286 random-effects parameters. The goodness of the final PK model was demonstrated by
287 GOF plots for observed versus model-predicted NVP concentrations. Overall data
288 spread around the line of identity suggesting adequate goodness of the PK model (see
289 supplementary Figure S3). For model evaluation, the final PK parameter estimates were
290 compared to the median and the 95% confidence interval obtained from the 1668
291 successful bootstrap runs (83.4%), see Table 2. For the fixed- and random-effects
292 parameters, the bootstrap medians were very similar to the original model parameter
293 values. All of them were within the 95% confidence interval, indicating an accurate and
294 precise description of the NVP data of both populations by the PK model.

295

296 *HIV-1 Transmission Risk under various NVP Single Dose Prophylaxis*
297 *Scenarios*

298 During the program for prevention of mother-to-child transmission in Uganda (28), NVP
299 was administered once to pregnant women during labor and once to newborns shortly
300 after delivery, with the aim of lowering the transmission probability of HIV-1 from mother-
301 to-child. The results and model-predicted HIV-1 transmission probabilities under the four
302 sd-NVP prophylaxis scenarios are illustrated in Figure 4 (A: no NVP prophylaxis, B:
303 single postpartum newborn NVP dose, C: single intrapartum maternal NVP dose and D:
304 intrapartum maternal NVP dose plus postpartum newborn NVP dose). The model-
305 predicted transmission risks agreed very well with published data from various trials (3,
306 19, 27, 34, 38, 49-50, 60). Without prophylaxis, the estimated HIV-1 transmission
307 probability after 2 years was $35.8 \pm 2.9\%$ (Figure 4 A). A single postpartum newborn
308 dose reduced the transmission probability to $31.6 \pm 2.7\%$ (Figure 4 B) whereas a single
309 intrapartum maternal dose lowered the transmission probability substantially to 19.1
310 $\pm 2.2\%$ (Figure 4 C). The combination of maternal and newborn doses reduced the
311 transmission probability to $19.7 \pm 2.2\%$ (Figure 4 D), which is insignificantly different
312 from a single maternal dose alone. The intrapartum infection risk (typically assessed
313 2 weeks after birth) was $18 \pm 2.4\%$, $12.3 \pm 2\%$, $0.1 \pm 0.1\%$ and $0.1 \pm 0.1\%$ for the four
314 investigated regimens. From the shape of the curves in Figure 4 A-D it can also be seen
315 that the subsequent risk of HIV-1 transmission (mainly through breastfeeding) is highest
316 during the first 200 days after birth.

317 *Mechanism of Prevention of Intrapartum HIV-1 Transmission by Maternal*
318 *NVP Prophylaxis*

319 Our data indicate that maternal NVP single dose alone decreases the transmission risk
320 of HIV-1 substantially compared to newborn NVP dose alone (compare Figure 4 B with
321 Figure 4 C). Hence, we elucidated the mechanisms by which the maternal NVP dose
322 lowers HIV-1 transmission probabilities.

323 The dynamics of viral load decay in the HIV-1 infected mother after the maternal NVP
324 dose are shown in Figure 5 A. The viral load declined by less than a factor of two during
325 the first 30 h after single dose NVP. However, the newborn was born within a range of
326 0.3-24.8 h, median: 5.1 h (28) (dashed horizontal bar and the open circle in Figure 5 A),
327 indicating that the maternal dose had little or no effect on the number of virions that
328 come in contact with the newborn during intrapartum virus challenge.

329 In Figure 5 B the concentrations of NVP in a representative newborn from the PK
330 investigation are depicted at the time of delivery (intrapartum challenge). Since NVP is
331 known to cross the placenta (35, 37) and this process was quantified by us (PCL, PCM,
332 see above), a fraction of the maternal NVP concentration was present in the newborn at
333 the time of delivery, where it is able to prevent HIV from infecting cells (Figure 5 B) by
334 lowering the infection rate β (see *Methods* section).

335 *Predictors for Selection and Persistence of NVP-resistant HIV-1 Strains in*
336 *Mothers after NVP Single Dose Administration*

337 Previous studies reported that a single dose of NVP can already select drug resistant
338 viral strains in the HIV-1 infected mothers (17, 22), compromising subsequent maternal
339 treatment success (8, 25, 29) and potentially promoting the transmission of NVP

340 resistant strains to the child during subsequent breastfeeding. We wanted to assess
341 predictors for the selection of drug resistant strains in HIV-1 infected mothers, which
342 might subsequently lead to the transmission of resistant virus to the breastfed child. Our
343 model predictions revealed a strong correlation between the individual half-life of NVP in
344 mothers and the duration in which NVP-resistant strains dominated the viral population
345 in the HIV-1 infected mothers after a single intrapartum maternal NVP dose, see Figure
346 6 A (spearman's rank correlation coefficient $r_s^2 = 0.98$). The model-predicted dynamics of
347 resistance appearance and fading for some representative mothers are shown in Figure
348 6 B-E. Our model predictions indicate that depending on the individual pharmacokinetics
349 of NVP, NVP-resistant strains become selected and might subsequently dominate the
350 virus population until NVP will be eliminated and resistant virus will be outgrown by the
351 wild type (see Figure 6 B-E) once again. This has important implications on the
352 probability that resistance is transmitted from mother-to-child and on the success of
353 subsequent extended newborn NVP prophylaxis.

354 In supplementary text S2, we derived equations that clarify the relation between
355 individual NVP concentrations C and resistance selection. Using these equations, it is
356 possible to compute the minimum NVP concentration that favors the selection of a
357 resistant strain over the wild type virus. For mutant K103N, Y181C and the double
358 mutant (K103N/Y181C) the determined minimum concentrations that favor their
359 selection are 6.56 ng/mL, 17.7 ng/mL and 21.6 ng/mL, respectively, based on the
360 phenotypic parameters used in this work ($IC_{50}(wt)$, $s(res)$ and $h(i)$).

361 This indicates, that single-point mutations are already selected at concentrations below
362 the IC_{50} value of the wild type (22 ng/mL (47)), which can persist in the plasma of the
363 mother for several weeks after sd-NVP, depending on the individual pharmacokinetic

364 NVP concentration-time profile. More importantly, if transmission of HIV-1 from mother-
365 to-child occurs during the particular time frame when the resistant virus dominates, it will
366 likely involve resistant virus and therefore lead to resistance spread.

367 *Extended NVP Prophylaxis Strategies to Prevent HIV-1 Transmission via* 368 *Breastfeeding*

369 We explored the impact of extended newborn NVP prophylaxis on HIV-1 transmission
370 risk in order to evaluate whether these may, similar to pre-exposure viral prophylaxis,
371 decrease the probability that viral challenges lead to infection in breastfed infants. In
372 addition to the maternal dose, we analyzed the impact of 6 weeks (SWEN-study, [17-
373 18]), 14-, 21 weeks, 6 month (HPTN 046-study, [19]), 52-, 78- or 104 weeks of extended
374 newborn NVP 2 mg/kg dosing, on the transmission risk of HIV-1. The predictions for
375 6 weeks and 6 months extended newborn NVP are displayed in Figure 7 A and Figure
376 7 B, respectively, together with clinical data from the SWEN-study (3, 38) (6 weeks
377 extended NVP) and the HPTN 046 trial (10) (6 month extended NVP). The agreement
378 between predicted- and observed transmission probabilities was very good. The
379 cumulative HIV-1 transmission risk 2 years postpartum, in the case of 6-, 14-, 21 weeks,
380 6 month, 52-, 78- or 104 weeks of extended NVP dosing were 19.6% \pm 2.1%, 15.8%
381 \pm 1.9%, 15.5% \pm 1.9%, 15.8% \pm 1.9%, 11.8% \pm 1.6%, 10.4% \pm 1.4% and 8.5% \pm 1.1%,
382 respectively (see Figure 7 C): All except the 6 weeks extended NVP regimen
383 significantly reduced HIV-1 transmission during 2 years postpartum, compared to
384 intrapartum single dose maternal/newborn NVP (cross-tab χ^2 test, $p < 0.05$,
385 respectively). Notably, the 52- and 104 weeks regimens reduced the risk of transmission
386 by further 50% and 60% compared to intrapartum maternal/newborn NVP dose alone.

387 The reduction of HIV-1 transmission per week extended NVP was 0.02%, 0.28%, 0.20%,
388 0.14%, 0.15%, 0.12% and 0.10% for the 6-, 14-, 21 weeks, 6 months, 52-, 78- or
389 104 weeks regimens, respectively.

390 *Probability of Transmitting Resistant Virus during Extended NVP* 391 *Prophylaxis*

392 The proportion of infections with NVP-resistant virus among the newborns that became
393 infected $P(\text{res. l inf.})_{0-2y}$ for the entire evaluation period (2 years postpartum) was 23.1%,
394 25.7%, 33.0%, 30.3%, 49.3%, 60.0% and 100%, respectively, in the 6-, 14-, 21 weeks,
395 6 months, 52-, 78- or 104 weeks extended NVP regimens (it was 22.3% in the single
396 dose intrapartum maternal- plus postpartum newborn regimen), neglecting intrauterine
397 infection. The proportion of infections with NVP-resistant virus among the infected
398 newborns during weeks 0-6 and > 6 weeks postpartum were $P(\text{res. l inf.})_{0-6w} = 100\%$ and
399 $P(\text{res. l inf.})_{>6w} = 16.9\%$, respectively in the 6 weeks extended NVP regimen, which is in
400 good agreement with published data from the SWEN study ($P(\text{res. l inf.})_{0-6w} = 92\%$ and
401 $P(\text{res. l inf.})_{>6w} = 15\%$, respectively (36)). For a single maternal and newborn NVP dose,
402 the conditional probabilities were $P(\text{res. l inf.})_{0-6w} = 73.9\%$ and $P(\text{res. l inf.})_{>6w} = 12.8\%$,
403 which overestimates the transmission of resistant strains during week 0-6, but agrees
404 well published data on resistance transmission after 6 weeks ($P(\text{res. l inf.})_{0-6w} = 38\%$ and
405 $P(\text{res. l inf.})_{>6w} = 15\%$, respectively (36)). The total number of infants infected with
406 resistant virus during the breastfeeding period $P(\text{res. l inf.}) \cdot P(\text{inf.})$ was not significantly
407 different in any extended newborn NVP regimen (3.38%, 2.77%, 3.47%, 3.27%, 3.35%,
408 3.24% and 3.50%, respectively in the 6-, 14-, 21 weeks, 6 month, 52-, 78- or 104 weeks

409 regimens, neglecting intrauterine infection) and was very similar to the single dose
410 intrapartum maternal- plus postpartum newborn regimen (3.28%). Our results indicated
411 that extended NVP only allows infection with resistant virus during the duration of its
412 administration. Our predictions also indicated that all infections with resistant virus
413 occurred before 200 days postpartum in agreement with resistance domination in the
414 breastfeeding mothers (shown in Figure 6).

415 **Discussion**

416 Short-course NVP prophylaxis is still widely used in resource-constrained settings to
417 prevent mother-to-child transmission of HIV-1. Since pregnant women and their
418 newborns represent particular subpopulations, plasma of mothers and newborns were
419 sampled for PK investigation during a Ugandan program for the prevention of mother-to-
420 child transmission, which comprised sd-NVP to pregnant women and newborns each.
421 For PK analysis of the NVP data, a combined population PK model was developed and
422 subsequently incorporated into pharmacodynamic (PD) investigations.

423 We found, in agreement with similar studies (4, 11, 13, 28), that a one-compartment
424 model with first-order absorption and elimination processes was sufficient to describe
425 the pharmacokinetics of NVP in pregnant women/mothers and newborns. Based on our
426 previously published separated PK models for pregnant women/mothers and newborns
427 (28), we developed a combined PK model in the present work that simultaneously
428 analyzed the NVP concentrations of pregnant women/mothers and newborns. Before
429 delivery, the PK model constituted the structure of a two-compartment model, where the
430 central- and peripheral compartments were linked to the pregnant women/mothers and
431 the fetus, respectively. Utilizing this model structure, we were able to estimate the

432 plasma/placenta transfer of NVP as newborns presented measureable NVP plasma
433 concentrations before receiving their own NVP dose. After delivery the combined PK
434 model for pregnant women/mothers and fetus was separated into two one-compartment
435 models for mothers and newborns, respectively. All PK parameters were precisely
436 estimated as shown by small relative standard errors. The estimated relative volume of
437 distribution in mothers was very high ($V_2/F = 91$ L) and in excellent agreement with
438 previously published values (range: 77-106 L) (4, 13, 26, 37). Maternal NVP elimination
439 capacity was low ($CL_1/F = 1.22$ L/h) and within the range of previously published values
440 (1.23-1.42 L/h) (4, 11, 37). The calculated half-life of NVP in mothers was 52 h, being
441 also within the range of previously published values 43-61 h (4, 11, 37). The half-life in
442 newborns (66 h) was slightly longer than the published value of 47 h (37), but
443 considerably shorter than the value of 110 h, observed in (4). However, in the previous
444 study (4), newborn plasma was only sampled over a very short interval (0-50 h),
445 whereas data in our investigation was sampled over a considerably longer period of time
446 (0-420 h), allowing to more accurately determine the elimination of NVP in newborns.
447 The evaluation of the final combined PK model by GOF plots and VPC demonstrated
448 appropriateness and sufficient predictive performance. Hence, the PK model could be
449 used as an input for further PD investigations. In order to simultaneously analyze the
450 impact of NVP pharmacokinetics on HIV-1 acquisition in the newborn, we developed a
451 PK-coupled stochastic HIV-1 dynamics model. Models for HIV-1 dynamics in
452 asymptotically infected individuals are rather established (reviewed in (40)). Few *in*
453 *silico* studies have linked viral dynamics to pharmacokinetics (15, 21, 43), modeled the
454 impact of pharmacokinetics on the emergence of drug resistance (54), or considered the
455 dynamics of HIV-1 infection (51-52). However, all these aspects, which concurrently

456 occur *in vivo*, have, to the authors' knowledge, never been addressed simultaneously by
457 mathematical modeling. In this study, we combined all these aspects in a single model.
458 Furthermore, our model considers many aspects of child growth, immune system
459 development and the characteristics of viral challenge during delivery and breastfeeding,
460 which have been validated with external data (see Figure S1, supplementary material).
461 Although no parameter adjustments for the HIV-1 dynamics model have been performed,
462 model-predicted HIV-1 transmission rates under various NVP-based treatment scenarios
463 were in excellent agreement with data from nine independent studies (see Figure 4 and
464 Figure 7), confirming the validity of the chosen approach.

465 Throughout this work, a reduced virus dynamics model was used, which is suited to
466 accurately predict viral load decay in HIV-1 infected individuals following single dose
467 administration of NVP and to predict the subsequent risk of child infection. In the case of
468 multiple dose maternal drug administration, we recommend to use a model that can
469 capture all phases of viral load decline, e.g. (55). In the present analysis we did not
470 focus on viral load dynamics after the infection of the child, but rather focused on the
471 infection risk (respective simulations were stopped, if newborn infection occurred). For
472 accurately analyzing viral load dynamics in infected children, we also recommend to use
473 more elaborated viral dynamics models, e.g. (55).

474 Our predictions indicated a significant impact of maternal NVP administration on the
475 reduction of HIV-1 transmission to the newborn (see Figure 4 C). An analysis of the
476 HIV-1 dynamics in the pregnant women between the period of NVP administration and
477 delivery indicated that the effect of maternal NVP on intrapartum transmission was not
478 due to a reduction in the number of virus particles potentially coming into contact with
479 the newborn during delivery, since viral load decayed only by less than a factor of two

480 during the first 30 h after NVP administration (see Figure 5 A). This model-derived result
481 is confirmed by clinically observed delays in virus load decline for NVP monotherapy
482 (24-48 h (20)). Likewise, delays in the onset of viral decay have been observed in the
483 case of ritonavir monotherapy (~30 h (41)) and under highly active antiretroviral therapy
484 (HAART) (~18 h (31)). We therefore conclude that a maternal dose, administered at the
485 onset of labor, may hardly have an impact of the number of viruses that come into
486 contact with the newborn during delivery. Instead, the PK analysis coupled to the virus
487 dynamics model, revealed that the main effect of the maternal dose is to provide
488 potentially protective NVP concentrations via *trans-placenta* transport to the newborn at
489 the moment of virus contact during delivery (see Figure 5 B), subsequently preventing
490 HIV-1 infection. These finding were confirmed by rapid NVP exchange through the
491 placenta (as indicated by the exchange parameters PCL, PCM in Table 2 and the almost
492 identical time points of maximum concentrations (t_{\max}) values in maternal and newborn
493 plasma and cord blood (4)). This mechanism of HIV-1 transmission prevention provided
494 by the maternal single dosing is highly similar to a pre-exposure prophylaxis, which has
495 recently demonstrated high potential in reducing HIV-1 transmission in the context of
496 sexual HIV-1 transmission (18). This particular mechanism of HIV-1 prevention by
497 maternal sd-NVP has important implications for the timing of the maternal dose: Since
498 trans-placental exchange is rapid (4), the newborn's NVP concentrations during delivery
499 would offer maximal protective effect at t_{\max} (mother) of 3.5 h [range: 3.0-4.1 h]
500 (calculated from individual PK parameter estimates). While NVP is absorbed rapidly (9),
501 HIV-1 prevention by the maternal dose is likely suboptimal before t_{\max} (mother). The
502 protective effect however lasts for relatively long periods of time, since NVP is slowly

503 eliminated (4, 9, 28) (see also Table 2). This indicates that the maternal NVP
504 administration at the onset of labor might be most effective, if feasible.

505 A single dose of NVP can select drug resistant viral strains in the HIV-infected mothers
506 (17, 22) (see Figure 6) and lead to transmission of NVP resistant strains to the child (e.g.
507 via breastfeeding). Pooled estimates showed that 36% (19–76%) of women have
508 detectable NVP resistance mutations 6–8 weeks after exposure to a single dose of NVP
509 (2). Our model slightly overestimated resistance development in the mothers after
510 receiving a single intrapartum NVP dose (62% and 70% at week 8 if the detection limit
511 for resistance was 50% and 20% respectively). This overestimation can be partially
512 explained by the use of a simplified model of resistance development in our
513 computational study, which ignores the genetic background on which resistance
514 develops; e.g. if resistance develops on some viral strain, which is particularly unfit, then
515 the resistance is less likely to be selected, see parameter $s(\text{res})$ in Equation (7). Instead,
516 in order to reduce the complexity of our mathematical model (and to reduce the
517 computational cost), we assumed that all susceptible viral strains were as fit as the
518 wild type and therefore all viral strains that develop a particular mutation (K103N, Y181C
519 and K103N/Y181C) were only assigned a fitness loss that comes from the resistance
520 mutation and not from the genetic background of the founder strain. In future, more
521 realistic and computationally feasible solutions for this problem should be developed.
522 Nevertheless, our estimates of resistance transmission to the newborns/infants were in
523 good agreement with clinical data from the SWEN study (36).

524 Our model predictions suggested a correlation between the individual half-life of NVP in
525 mothers and the duration in which NVP-resistant strains dominated the viral population
526 in the HIV-1 infected mothers after a single intrapartum maternal NVP dose. Selection of

527 resistant strains could be explained mathematically (see supplementary material) and
528 minimum concentrations for the selection of NVP-resistant strains were derived.
529 Combining the pharmacokinetic analysis of individual pharmacokinetics with the model
530 of HIV-1 dynamics and transmission, we predicted that transmission of NVP-resistant
531 strains would occur during the first 200 days after single dose maternal NVP, in line with
532 the time frame in which resistant strains likely dominate the viral population (Figure 6).
533 Figure 6 A-B suggest that NVP resistance might not become selected in mothers after
534 single dose administration, if individual NVP elimination is fast enough (short NVP half-
535 life). This indicates that resistance selection and subsequent resistance transmission to
536 the child via breastfeeding could be reduced if drugs were administered to the mothers,
537 which, in contrast to NVP, exhibit a very short half-life (e.g. zidovudine). However, we
538 also showed that NVP effectively prevents intrapartum HIV transmission by being
539 transferred across the placenta to the child, so that any drug which might replace
540 maternal single dose NVP should also be able to cross the placenta in order to
541 effectively protect the child from infection during the birth process (see Figure 5). Adding
542 drugs to the maternal sd-NVP is another effective approach to reduce resistance
543 selection in the HIV-1 infected mothers and to further lower intrapartum transmission
544 rates (5, 7, 33), potentially by increasing the genetic barrier to resistance selection. A
545 thorough understanding of the underlying mechanisms, however, is still lacking and
546 mathematical models including combinations of drugs for elucidation remain to be
547 developed in future.

548 Currently, two main strategies are pursued in order to reduce subsequent HIV-1
549 transmission via breastfeeding: (i) maternal ART or (ii) extended newborn NVP
550 prophylaxis. Maternal ART has been shown to reduce HIV-1 transmission via

551 breastfeeding, by lowering maternal viral load to less than 400 copies per mL (14, 46),
552 but long-term drug treatment might not be available in resource-limited settings.
553 Extended newborn NVP administration has been suggested to reduce the transmission
554 risk of HIV-1 by postpartum breastfeeding and might be the regimen of choice in
555 extremely resource-limited settings for reasons of cost-effectiveness compared to
556 maternal ART (57). In Figure 7, we analyzed the impact of 6-, 14-, 21 weeks, 6 month,
557 52-, 78- or 104 weeks extended newborn NVP on the transmission risk of HIV-1. Our
558 data agrees very well with published data from the SWEN-study (3, 38) (6 weeks
559 extended NVP) and the HPTN 049-study (10) (6 month extended NVP). Although a
560 reduction of the HIV-1 transmission risk at 1 year postpartum was reported in the
561 SWEN-study (6 weeks extended NVP), this reduction was not significantly different from
562 single dose intrapartum maternal and newborn NVP dose alone (13.9% vs. 15.4%, $p =$
563 0.33; including 5% intrauterine transmission probability) (38). Our results support this
564 finding: The estimated transmission probability 1 year postpartum was $15.3 \pm 1.9\%$ and
565 $16.8 \pm 2\%$ $p = 0.28$ (including 5% intrauterine transmission probability), respectively for 6
566 weeks extended NVP and single dose intrapartum maternal and newborn NVP dose. At
567 2 years postpartum a significant reduction in the HIV-1 transmission could be achieved
568 for all investigated extended NVP regimens, except the 6 weeks extended NVP regimen,
569 in comparison to single dose intrapartum maternal and newborn NVP dose alone. The
570 cost-effectiveness however decreases with increasing length of extended NVP as
571 reflected by the reduction of HIV-1 transmission per week of extended newborn NVP.
572 This indicates that although substantial further decrease of HIV-1 transmission could be
573 achieved by extended NVP regimens which cover most of the breastfeeding period,
574 shorter periods of extended NVP might be more feasible in (extremely) resource-limited

575 settings with regard to cost-effectiveness. Our estimates of resistance transmission to
576 the newborns were in good agreement with clinical data from the SWEN-study (36).
577 Overall, our results indicated an increase in the *proportion* of infections with resistant
578 virus for longer durations of extended NVP prophylaxis. However, the *total number* of
579 newborns who become infected with resistant virus was not increased by any of the
580 extended NVP prophylaxis regimens compared to NVP single dose; mainly because
581 extended NVP simultaneously minimizes the transmission probability.

582 Summarized, we have developed a coupled *in vitro/in vivo* pharmacokinetic-
583 pharmacodynamic model to assess the effects of distinct NVP prophylaxis regimens on
584 the prevention of mother-to-child transmission of HIV-1 and resistance formation. Our
585 model shows very good predictive performance compared to data from clinical studies.
586 The model may be adapted to predict the outcome of other drug interventions and could
587 therefore be used as a supportive tool to improve HIV-1 prevention, maximize cost-
588 effectiveness and reduce risk of resistance selection when novel studies are planned.

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595 **References**

- 596 1. *Alfonsi, A., E. Cances, Turinici G., Ventura B., and W. Huisinga. 2005. Exact*
597 *simulation of hybrid stochastic and deterministic models for biochemical systems.*
598 *ESAIM Proc 14:1-23.*
- 599 2. *Arrive, E., M.-L. Newell, D. K. Ekouevi, M.-L. Chaix, R. Thiebaut, B. Masquelier, V.*
600 *Leroy, P. V. d. Perre, C. Rouzioux, and F. Dabis. 2007. Prevalence of resistance to*
601 *nevirapine in mothers and children after single-dose exposure to prevent vertical*
602 *transmission of HIV-1: a meta-analysis. Int J Epidemiol 36:1009-1021.*
- 603 3. *Bedri, A., B. Gudetta, A. Isehak, S. Kumbi, S. Lulseged, Y. Mengistu, A. V. Bhore, R.*
604 *Bhosale, V. Varadhrajan, N. Gupte, J. Sastry, N. Suryavanshi, S. Tripathy, F. Mmiro,*
605 *M. Mubiru, C. Onyango, A. Taylor, P. Musoke, C. Nakabiito, A. Abashawl, R. Adamu,*
606 *G. Antelman, R. C. Bollinger, P. Bright, M. A. Chaudhary, J. Coberly, L. Guay, M. G.*
607 *Fowler, A. Gupta, E. Hassen, J. B. Jackson, L. H. Moulton, U. Nayak, S. B. Omer, L.*
608 *Propper, M. Ram, V. Rexroad, A. J. Ruff, A. Shankar, and S. Zwierski. 2008. Extended-*
609 *dose nevirapine to 6 weeks of age for infants to prevent HIV transmission via*
610 *breastfeeding in Ethiopia, India, and Uganda: an analysis of three randomised*
611 *controlled trials. Lancet 372:300-313.*
- 612 4. *Benaboud, S., D. K. Ekouevi, S. Urien, E. Rey, E. Arrive, S. Blanche, G. Gray, K. L.*
613 *Sim, D. Avit, J. McIntyre, E. Nerrienet, F. Dabis, J. M. Treluyer, and D. Hirt. 2010.*
614 *Population Pharmacokinetics of Nevirapine in HIV-1 infected Pregnant Women and*
615 *their neonates (ANRS 12109). Antimicrob Agents Chemother.*
- 616 5. *Chaix, M. L., D. K. Ekouevi, F. Rouet, B. Tonwe-Gold, I. Viho, L. Bequet, G. Peytavin,*
617 *H. Toure, H. Menan, V. Leroy, F. Dabis, and C. Rouzioux. 2006. Low risk of*
618 *nevirapine resistance mutations in the prevention of mother-to-child transmission of*
619 *HIV-1: Agence Nationale de Recherches sur le SIDA Ditrane Plus, Abidjan, Cote*
620 *d'Ivoire. J Infect Dis 193:482-487.*
- 621 6. *Chasela, C. S., M. G. Hudgens, D. J. Jamieson, D. Kayira, M. C. Hosseinipour, A. P.*
622 *Kourtis, F. Martinson, G. Tegha, R. J. Knight, Y. I. Ahmed, D. D. Kamwendo, I. F.*
623 *Hoffman, S. R. Ellington, Z. Kacheche, A. Soko, J. B. Wiener, S. A. Fiscus, P.*
624 *Kazembe, I. A. Mofolo, M. Chigwenembe, D. S. Sichali, and C. M. van der Horst. 2010.*
625 *Maternal or infant antiretroviral drugs to reduce HIV-1 transmission. N Engl J Med*
626 *362:2271-2281.*
- 627 7. *Chi, B. H., M. Sinkala, F. Mbewe, R. A. Cantrell, G. Kruse, N. Chintu, G. M.*
628 *Aldrovandi, E. M. Stringer, C. Kankasa, J. T. Safrit, and J. S. Stringer. 2007. Single-*
629 *dose tenofovir and emtricitabine for reduction of viral resistance to non-nucleoside*
630 *reverse transcriptase inhibitor drugs in women given intrapartum nevirapine for*
631 *perinatal HIV prevention: an open-label randomised trial. Lancet 370:1698-1705.*
- 632 8. *Coffie, P. A., D. K. Ekouevi, M. L. Chaix, B. Tonwe-Gold, A. B. Clarisse, R. Becquet, I.*
633 *Viho, T. N'Dri-Yoman, V. Leroy, E. J. Abrams, C. Rouzioux, and F. Dabis. 2008.*

- 634 *Maternal 12-month response to antiretroviral therapy following prevention of mother-*
635 *to-child transmission of HIV type 1, Ivory Coast, 2003-2006. Clin Infect Dis 46:611-621.*
- 636 9. Cooper, C. L., and R. P. van Heeswijk. 2007. *Once-daily nevirapine dosing: a*
637 *pharmacokinetics, efficacy and safety review. HIV Med 8:1-7.*
- 638 10. Coovadia, H., E. Brown, Y. Maldonado, L. Mofenson, D. Moodley, P. Musoke, M. G.
639 Fowler, K. Manji, K. Geoge, S. Zwierski, and H. P. Team. 2011. *HPTN: Efficacy of*
640 *Extended Daily Infant NVP through Age 6 Month Compared to 6 Weeks for Postnatal*
641 *PMTCT of HIV through Breastfeeding Conference on Retroviruses and Opportunistic*
642 *Infections (<http://www.retroconference.org/2011/Abstracts/42412.htm>).*
- 643 11. Cressey, T. R., G. Jourdain, M. J. Lallemand, S. Kunkeaw, J. B. Jackson, P. Musoke, E.
644 Capparelli, and M. Mirochnick. 2005. *Persistence of nevirapine exposure during the*
645 *postpartum period after intrapartum single-dose nevirapine in addition to zidovudine*
646 *prophylaxis for the prevention of mother-to-child transmission of HIV-1. J Acquir*
647 *Immune Defic Syndr 38:283-288.*
- 648 12. De Cock, K. M., M. G. Fowler, E. Mercier, I. de Vincenzi, J. Saba, E. Hoff, D. J.
649 Alnwick, M. Rogers, and N. Shaffer. 2000. *Prevention of mother-to-child HIV*
650 *transmission in resource-poor countries: translating research into policy and practice.*
651 *JAMA 283:1175-1182.*
- 652 13. de Maat, M. M., A. D. Huitema, J. W. Mulder, P. L. Meenhorst, E. C. van Gorp, and J.
653 H. Beijnen. 2002. *Population pharmacokinetics of nevirapine in an unselected cohort*
654 *of HIV-1-infected individuals. Br J Clin Pharmacol 54:378-385.*
- 655 14. de Vincenzi, I. 2011. *Triple antiretroviral compared with zidovudine and single-dose*
656 *nevirapine prophylaxis during pregnancy and breastfeeding for prevention of mother-*
657 *to-child transmission of HIV-1 (Kesho Bora study): a randomised controlled trial.*
658 *Lancet Infect Dis 11:171-180.*
- 659 15. Dixit, N. M., and A. S. Perelson. 2004. *Complex patterns of viral load decay under*
660 *antiretroviral therapy: influence of pharmacokinetics and intracellular delay. J Theor*
661 *Biol 226:95-109.*
- 662 16. Elsherbiny, D., K. Cohen, B. Jansson, P. Smith, H. McIlleron, and U. S. Simonsson.
663 2009. *Population pharmacokinetics of nevirapine in combination with rifampicin-based*
664 *short course chemotherapy in HIV- and tuberculosis-infected South African patients.*
665 *Eur J Clin Pharmacol 65:71-80.*
- 666 17. Eshleman, S. H., M. Mracna, L. A. Guay, M. Deseve, S. Cunningham, M. Mirochnick,
667 P. Musoke, T. Fleming, M. Glenn Fowler, L. M. Mofenson, F. Mmiro, and J. B.
668 Jackson. 2001. *Selection and fading of resistance mutations in women and infants*
669 *receiving nevirapine to prevent HIV-1 vertical transmission (HIVNET 012). AIDS*
670 *15:1951-1957.*
- 671 18. Grant, R. M., J. R. Lama, P. L. Anderson, V. McMahan, A. Y. Liu, L. Vargas, P.
672 Goicochea, M. Casapia, J. V. Guanira-Carranza, M. E. Ramirez-Cardich, O. Montoya-

- 673 *Herrera, T. Fernandez, V. G. Veloso, S. P. Buchbinder, S. Chariyalertsak, M.*
674 *Schechter, L. G. Bekker, K. H. Mayer, E. G. Kallas, K. R. Amico, K. Mulligan, L. R.*
675 *Bushman, R. J. Hance, C. Ganoza, P. Defechereux, B. Postle, F. Wang, J. J.*
676 *McConnell, J. H. Zheng, J. Lee, J. F. Rooney, H. S. Jaffe, A. I. Martinez, D. N. Burns,*
677 *and D. V. Glidden. 2010. Preexposure Chemoprophylaxis for HIV Prevention in Men*
678 *Who Have Sex with Men. N Engl J Med.*
- 679 19. *Guay, L. A., P. Musoke, T. Fleming, D. Bagenda, M. Allen, C. Nakabiito, J. Sherman,*
680 *P. Bakaki, C. Ducar, M. Deseyve, L. Emel, M. Mirochnick, M. G. Fowler, L. Mofenson,*
681 *P. Miotti, K. Dransfield, D. Bray, F. Mmiro, and J. B. Jackson. 1999. Intrapartum and*
682 *neonatal single-dose nevirapine compared with zidovudine for prevention of mother-to-*
683 *child transmission of HIV-1 in Kampala, Uganda: HIVNET 012 randomised trial.*
684 *Lancet 354:795-802.*
- 685 20. *Havlir, D. V., S. Eastman, A. Gamst, and D. D. Richman. 1996. Nevirapine-resistant*
686 *human immunodeficiency virus: kinetics of replication and estimated prevalence in*
687 *untreated patients. Journal of virology 70:7894-7899.*
- 688 21. *Huang, Y., S. L. Rosenkranz, and H. Wu. 2003. Modeling HIV dynamics and antiviral*
689 *response with consideration of time-varying drug exposures, adherence and phenotypic*
690 *sensitivity. Math Biosci 184:165-186.*
- 691 22. *Jackson, J. B., G. Becker-Pergola, L. A. Guay, P. Musoke, M. Mracna, M. G. Fowler, L.*
692 *M. Mofenson, M. Mirochnick, F. Mmiro, and S. H. Eshleman. 2000. Identification of*
693 *the K103N resistance mutation in Ugandan women receiving nevirapine to prevent*
694 *HIV-1 vertical transmission. AIDS 14:F111-115.*
- 695 23. *Jackson, J. B., P. Musoke, T. Fleming, L. A. Guay, D. Bagenda, M. Allen, C. Nakabiito,*
696 *J. Sherman, P. Bakaki, M. Owor, C. Ducar, M. Deseyve, A. Mwatha, L. Emel, C.*
697 *Duefield, M. Mirochnick, M. G. Fowler, L. Mofenson, P. Miotti, M. Gigliotti, D. Bray,*
698 *and F. Mmiro. 2003. Intrapartum and neonatal single-dose nevirapine compared with*
699 *zidovudine for prevention of mother-to-child transmission of HIV-1 in Kampala,*
700 *Uganda: 18-month follow-up of the HIVNET 012 randomised trial. Lancet 362:859-*
701 *868.*
- 702 24. *John-Stewart, G., D. Mbori-Ngacha, R. Ekpini, E. N. Janoff, J. Nkengasong, J. S.*
703 *Read, P. Van de Perre, and M. L. Newell. 2004. Breast-feeding and Transmission of*
704 *HIV-1. J Acquir Immune Defic Syndr 35:196-202.*
- 705 25. *Jourdain, G., N. Ngo-Giang-Huong, S. Le Coeur, C. Bowonwatanuwong, P. Kantipong,*
706 *P. Leechanachai, S. Ariyadej, P. Leenasirimakul, S. Hammer, and M. Lallemand. 2004.*
707 *Intrapartum exposure to nevirapine and subsequent maternal responses to nevirapine-*
708 *based antiretroviral therapy. N Engl J Med 351:229-240.*
- 709 26. *Kappelhoff, B. S., F. van Leth, T. R. MacGregor, J. Lange, J. H. Beijnen, and A. D.*
710 *Huitema. 2005. Nevirapine and efavirenz pharmacokinetics and covariate analysis in*
711 *the 2NN study. Antivir Ther 10:145-155.*

- 712 27. *Kumwenda, N. I., D. R. Hoover, L. M. Mofenson, M. C. Thigpen, G. Kafulafula, Q. Li,*
713 *L. Mipando, K. Nkanaunena, T. Mebrahtu, M. Bulterys, M. G. Fowler, and T. E. Taha.*
714 *2008. Extended antiretroviral prophylaxis to reduce breast-milk HIV-1 transmission. N*
715 *Engl J Med 359:119-129.*
- 716 28. *Kunz, A., M. Frank, K. Mugenyi, R. Kabasinguzi, A. Weidenhammer, M. Kurowski, C.*
717 *Kloft, and G. Harms. 2009. Persistence of nevirapine in breast milk and plasma of*
718 *mothers and their children after single-dose administration. J Antimicrob Chemother*
719 *63:170-177.*
- 720 29. *Lockman, S., R. L. Shapiro, L. M. Smeaton, C. Wester, I. Thior, L. Stevens, F. Chand,*
721 *J. Makhema, C. Moffat, A. Asmelash, P. Ndase, P. Arimi, E. van Widenfelt, L. Mazhani,*
722 *V. Novitsky, S. Lagakos, and M. Essex. 2007. Response to antiretroviral therapy after a*
723 *single, peripartum dose of nevirapine. N Engl J Med 356:135-147.*
- 724 30. *Mansky, L. M., and H. M. Temin. 1995. Lower in vivo mutation rate of human*
725 *immunodeficiency virus type 1 than that predicted from the fidelity of purified reverse*
726 *transcriptase. Journal of virology 69:5087-5094.*
- 727 31. *Markowitz, M., M. Louie, A. Hurley, E. Sun, M. Di Mascio, A. S. Perelson, and D. D.*
728 *Ho. 2003. A novel antiviral intervention results in more accurate assessment of human*
729 *immunodeficiency virus type 1 replication dynamics and T-cell decay in vivo. Journal*
730 *of virology 77:5037-5038.*
- 731 32. *Martinez-Picado, J., and M. A. Martinez. 2008. HIV-1 reverse transcriptase inhibitor*
732 *resistance mutations and fitness: a view from the clinic and ex vivo. Virus Res 134:104-*
733 *123.*
- 734 33. *McIntyre, J. A., M. Hopley, D. Moodley, M. Eklund, G. E. Gray, D. B. Hall, P.*
735 *Robinson, D. Mayers, and N. A. Martinson. 2009. Efficacy of short-course AZT plus*
736 *3TC to reduce nevirapine resistance in the prevention of mother-to-child HIV*
737 *transmission: a randomized clinical trial. PLoS Med 6:e1000172.*
- 738 34. *Miotti, P. G., T. E. Taha, N. I. Kumwenda, R. Broadhead, L. A. Mtimavalye, L. Van der*
739 *Hoeven, J. D. Chiphangwi, G. Liomba, and R. J. Biggar. 1999. HIV transmission*
740 *through breastfeeding: a study in Malawi. JAMA 282:744-749.*
- 741 35. *Mirochnick, M., T. Fenton, P. Gagnier, J. Pav, M. Gwynne, S. Siminski, R. S. Sperling,*
742 *K. Beckerman, E. Jimenez, R. Yogev, S. A. Spector, and J. L. Sullivan. 1998.*
743 *Pharmacokinetics of nevirapine in human immunodeficiency virus type 1-infected*
744 *pregnant women and their neonates. Pediatric AIDS Clinical Trials Group Protocol*
745 *250 Team. The Journal of infectious diseases 178:368-374.*
- 746 36. *Moorthy, A., A. Gupta, R. Bhosale, S. Tripathy, J. Sastry, S. Kulkarni, M. Thakar, R.*
747 *Bharadwaj, A. Kagal, A. V. Bhore, S. Patil, V. Kulkarni, V. Venkataramani, U.*
748 *Balasubramaniam, N. Suryavanshi, C. Ziemniak, N. Gupte, R. Bollinger, and D.*
749 *Persaud. 2009. Nevirapine resistance and breast-milk HIV transmission: effects of*
750 *single and extended-dose nevirapine prophylaxis in subtype C HIV-infected infants.*
751 *PLoS One 4:e4096.*

- 752 37. *Musoke, P., L. A. Guay, D. Bagenda, M. Mirochnick, C. Nakabiito, T. Fleming, T.*
753 *Elliott, S. Horton, K. Dransfield, J. W. Pav, A. Murarka, M. Allen, M. G. Fowler, L.*
754 *Mofenson, D. Hom, F. Mmiro, and J. B. Jackson. 1999. A phase I/II study of the safety*
755 *and pharmacokinetics of nevirapine in HIV-1-infected pregnant Ugandan women and*
756 *their neonates (HIVNET 006). AIDS 13:479-486.*
- 757 38. *Omer, S. B. 2011. Twelve-month follow-up of Six Week Extended Dose Nevirapine*
758 *randomized controlled trials: differential impact of extended-dose nevirapine on*
759 *mother-to-child transmission and infant death by maternal CD4 cell count. AIDS*
760 *25:767-776.*
- 761 39. *Palumbo, P., J. C. Lindsey, M. D. Hughes, M. F. Cotton, R. Bobat, T. Meyers, M.*
762 *Bwakura-Dangarembizi, B. H. Chi, P. Musoke, P. Kamthunzi, W. Schimana, L. Purdue,*
763 *S. H. Eshleman, E. J. Abrams, L. Millar, E. Petzold, L. M. Mofenson, P. Jean-Philippe,*
764 *and A. Violari. 2010. Antiretroviral treatment for children with peripartum nevirapine*
765 *exposure. N Engl J Med 363:1510-1520.*
- 766 40. *Perelson, A. S. 2002. Modelling viral and immune system dynamics. Nat Rev Immunol*
767 *2:28-36.*
- 768 41. *Perelson, A. S., A. U. Neumann, M. Markowitz, J. M. Leonard, and D. D. Ho. 1996.*
769 *HIV-1 dynamics in vivo: virion clearance rate, infected cell life-span, and viral*
770 *generation time. Science 271:1582-1586.*
- 771 42. *Petra_Study_Team. 2002. Efficacy of three short-course regimens of zidovudine and*
772 *lamivudine in preventing early and late transmission of HIV-1 from mother to child in*
773 *Tanzania, South Africa, and Uganda (Petra study): a randomised, double-blind,*
774 *placebo-controlled trial. Lancet 359:1178-1186.*
- 775 43. *Rosario, M. C., P. Jacqmin, P. Dorr, E. van der Ryst, and C. Hitchcock. 2005. A*
776 *pharmacokinetic-pharmacodynamic disease model to predict in vivo antiviral activity of*
777 *maraviroc. Clin Pharmacol Ther 78:508-519.*
- 778 44. *Sampah, M. E. S., L. Shen, B. L. Jilek, and R. F. Siliciano. 2011. Dose-response curve*
779 *slope is a missing dimension in the analysis of HIV-1 drug resistance. P Natl Acad Sci*
780 *USA 108:7613-7618.*
- 781 45. *Sedaghat, A. R., J. B. Dinoso, L. Shen, C. O. Wilke, and R. F. Siliciano. 2008. Decay*
782 *dynamics of HIV-1 depend on the inhibited stages of the viral life cycle. Proc Natl Acad*
783 *Sci U S A 105:4832-4837.*
- 784 46. *Shapiro, R. L., M. D. Hughes, A. Ogwu, D. Kitch, S. Lockman, C. Moffat, J. Makhema,*
785 *S. Moyo, I. Thior, K. McIntosh, E. van Widenfelt, J. Leidner, K. Powis, A. Asmelash, E.*
786 *Tumbare, S. Zwierski, U. Sharma, E. Handelsman, K. Mburu, O. Jayeoba, E. Moko, S.*
787 *Souda, E. Lubega, M. Akhtar, C. Wester, R. Tuomola, W. Snowden, M. Martinez-*
788 *Tristani, L. Mazhani, and M. Essex. 2010. Antiretroviral regimens in pregnancy and*
789 *breast-feeding in Botswana. N Engl J Med 362:2282-2294.*

- 790 47. Shen, L., S. Peterson, A. R. Sedaghat, M. A. McMahon, M. Callender, H. L. Zhang, Y.
791 Zhou, E. Pitt, K. S. Anderson, E. P. Acosta, and R. F. Siliciano. 2008. Dose-response
792 curve slope sets class-specific limits on inhibitory potential of anti-HIV drugs. *Nat Med*
793 *14*:762-766.
- 794 48. Stocker, H., G. Kruse, P. Kreckel, C. Herzmann, K. Arasteh, J. Claus, H. Jessen, C.
795 Cordes, B. Hintsche, F. Schlote, L. Schneider, and M. Kurowski. 2004. Nevirapine
796 significantly reduces the levels of racemic methadone and (R)-methadone in human
797 immunodeficiency virus-infected patients. *Antimicrob Agents Chemother* *48*:4148-4153.
- 798 49. Stringer, J. S., M. Sinkala, V. Chapman, E. P. Acosta, G. M. Aldrovandi, V. Mudenda,
799 J. P. Stout, R. L. Goldenberg, R. Kumwenda, and S. H. Vermund. 2003. Timing of the
800 maternal drug dose and risk of perinatal HIV transmission in the setting of intrapartum
801 and neonatal single-dose nevirapine. *AIDS* *17*:1659-1665.
- 802 50. Taha, T. E., N. I. Kumwenda, A. Gibbons, R. L. Broadhead, S. Fiscus, V. Lema, G.
803 Liomba, C. Nkhoma, P. G. Miotti, and D. R. Hoover. 2003. Short postexposure
804 prophylaxis in newborn babies to reduce mother-to-child transmission of HIV-1: NVAZ
805 randomised clinical trial. *Lancet* *362*:1171-1177.
- 806 51. Tuckwell, H. C., and E. Le Corfec. 1998. A stochastic model for early HIV-1 population
807 dynamics. *J Theor Biol* *195*:451-463.
- 808 52. Tuckwell, H. C., P. D. Shipman, and A. S. Perelson. 2008. The probability of HIV
809 infection in a new host and its reduction with microbicides. *Math Biosci* *214*:81-86.
- 810 53. UNAIDS. 2010. UNAIDS Report on the Global AIDS Epidemic (assessed April-26-
811 2011), http://www.unaids.org/globalreport/Global_report.htm.
- 812 54. von Kleist, M., and W. Huisinga. 2009. Pharmacokinetic-pharmacodynamic
813 relationship of NRTIs and its connection to viral escape: an example based on
814 zidovudine. *Eur J Pharm Sci* *36*:532-543.
- 815 55. von Kleist, M., S. Menz, and W. Huisinga. 2010. Drug-class specific impact of antivirals
816 on the reproductive capacity of HIV. *PLoS Comput Biol* *6*:e1000720.
- 817 56. Wei, X., S. K. Ghosh, M. E. Taylor, V. A. Johnson, E. A. Emini, P. Deutsch, J. D.
818 Lifson, S. Bonhoeffer, M. A. Nowak, B. H. Hahn, and et al. 1995. Viral dynamics in
819 human immunodeficiency virus type 1 infection. *Nature* *373*:117-122.
- 820 57. WHO. 2010. Antiretroviral drugs for treating pregnant women and preventing HIV
821 infection in infants: towards universal access (assessed April-26-2011),
822 <http://www.who.int/hiv/pub/mtct/antiretroviral2010/en/index.html>.
- 823 58. WHO. 2006. Antiretroviral drugs for treating pregnant women and preventing HIV
824 infection in infants: towards universal access (assessed April-26-2011),
825 <http://www.who.int/hiv/pub/mtct/antiretroviral/en/index.html>.

- 826 59. WHO. 2010. *Guidelines on HIV and infant feeding (assessed April-26-2011)*,
827 http://www.who.int/child_adolescent_health/documents/9789241599535/en/index.html.
- 828 60. Wiktor, S. Z., E. Ekpini, J. M. Karon, J. Nkengasong, C. Maurice, S. T. Severin, T. H.
829 Roels, M. K. Kouassi, E. M. Lackritz, I. M. Coulibaly, and A. E. Greenberg. 1999.
830 *Short-course oral zidovudine for prevention of mother-to-child transmission of HIV-1*
831 *in Abidjan, Cote d'Ivoire: a randomised trial. Lancet* 353:781-785.
- 832 61. Wilkinson, D. J. 2006. *Stochastic Modelling in Systems Biology*. Chapman & Hall/CRC,
833 Boca Raton.
- 834 62. Zhou, Y., H. Zhang, J. D. Siliciano, and R. F. Siliciano. 2005. *Kinetics of human*
835 *immunodeficiency virus type 1 decay following entry into resting CD4+ T cells. Journal*
836 *of virology* 79:2199-2210.

837 **Legends**

838 Figure 1: Final PK model of mother and newborn data. A: Observed NVP concentrations
839 in the plasma of HIV-1 infected pregnant women/mothers (filled diamonds) and in the
840 plasma of newborns (open triangles) sampled over three time periods: delivery, week 1
841 and week 2 after single dose of 200 mg NVP for pregnant women and 2 mg/kg NVP
842 administered to newborns (modified from (28)). B: Schematic structural model for the PK
843 of mothers and newborns. The absorption rate constant for oral dose of mothers and
844 newborns are K_A and K_{34} , respectively. V_2 describes the central volume of distribution
845 for maternal data. V_4 equals the volume of distribution of the peripheral compartment
846 (fetus/newborn compartment). Both compartments were linked by placenta clearance
847 (PCL) term before delivery. All dashed lines highlight time-dependent processes while
848 solid lines present continuous processes over the entire investigational period. The
849 partition coefficient fetus to pregnant women (PCM) denotes the ratio between NVP
850 concentrations in the fetus and maternal NVP concentrations before delivery and at
851 quasi steady state. NVP elimination from the central and the peripheral compartment
852 was described by CL_1 and CL_2 , respectively.

853 Figure 2: VPC of the final PK model of mother and newborn data. A and B: VPC of the
854 observed NVP concentrations in maternal plasma (black diamonds, A) and in newborn
855 plasma (open triangles, B) over time and 5th and 95th percentiles of model simulations
856 and model-simulated median (dashed- and solid lines).

857 Figure 3: Mathematical model of virus dynamics, mutation and transmission. A: Life-
858 cycle models of HIV-1 in mothers and newborns and their interconnection via
859 intrapartum- and breastfeeding challenge. Free virus can infect T-cells with infection rate

860 constant β , which encompasses all steps from target cell binding, fusion, to reverse
861 transcription. Early infected T-cells (after reverse transcription but prior to pro-virus
862 integration) become transformed into productively infected cells T2, after pro-virus
863 translocation into the nucleus and integration with rate k_T . Productively infected T-cells
864 T2 produce new virus V with rate N. Mutation occurs during the process of reverse
865 transcription (embodied in parameter β). NVP inhibits reverse transcription and therefore
866 affects parameter β in our model. All parameter values are listed in Table 1. Intrapartum
867 viral challenge occurs during delivery, whereas breastfeeding viral challenges occur
868 repeatedly after birth until the age of 2 years, according to the breastfeeding frequency
869 (Figure S2, supplementary material). B: Mutational graph showing the transition
870 probabilities $p_{j \rightarrow i}$ between the four virus strains (wild type *wt* and 3 mutants *K103N*,
871 *Y181C* and *K103N/Y181C*) considered here. C: Phenotypic attributes of the four
872 mutants. The extension of the bars to the right illustrates their IC_{50} value, whereas the
873 left extension indicates their fitness loss and the height of the bars indicates the slope
874 parameter. The IC_{50} -values were 22 ng/mL (47), 2168 ng/mL, 8671 ng/mL (44) and
875 >11500 ng/mL for *wt*, the *K103N*, the *Y181C* mutation and the double mutant
876 *K103N/Y181C*, respectively. The selective disadvantage s with respect to the wild type
877 was 12.5%, 40% and 52.5% for the *K103N*, the *Y181C* mutation and the double mutant
878 (32). The slope parameters were 1.55, 1.40, 1.15 and 1.0 for *wt*, the *K103N*, the *Y181C*
879 mutation and the double mutant *K103N/Y181C* (44, 47), respectively.

880 Figure 4: Cumulative HIV-1 transmission risk under various NVP single dose prophylaxis
881 strategies. Solid lines denote the Kaplan-Meier estimates of the model-predicted
882 cumulative probability of infection whereas light-grey areas represent the confidence
883 range for the model predictions. A: no NVP is given (upward- and downward pointing

884 triangles denote data from (60) and (34)); B: a single postpartum NVP dose (2 mg/kg) is
885 given to the newborn within 72 h after birth (squares denote data from (50)); C: a single
886 intrapartum NVP dose (200 mg) is given to the mother at the onset of labor; D: a single
887 intrapartum NVP dose (200 mg) and a single postpartum newborn dose (2 mg/kg) were
888 administered (crosses, open circles, diamonds, filled circles and plus signs denote data
889 from (3, 19, 27, 38, 49)). In all simulations, an intrauterine transmission probability of 5%
890 (12) was assumed.

891 Figure 5: A: Viral load (thick line) during the first 30 h in the plasma of HIV-1 infected
892 pregnant women/mothers after a single intrapartum dose NVP in relation to the time of
893 delivery (open circle denotes the median time of delivery, see *Methods* section and
894 dashed horizontal bar denotes the range). B: NVP concentration in a representative
895 newborn from the PK-investigation before- during- and after birth (solid line). The black
896 square and the black circle indicate the time of birth and the time of the newborn NVP
897 single oral dose in the representative newborn, respectively.

898 Figure 6: Predicted correlation between NVP elimination and persistence of NVP
899 resistance in HIV-1 positive mothers after a single dose of NVP. A: Correlation of
900 individual NVP half-life and predicted duration in which NVP resistance dominated the
901 viral population in mothers. B-E: Examples of resistance appearance and fading in
902 distinct, representative HIV-1 positive mothers after single dose NVP administration at
903 the onset of labor. Solid line: relative wild type abundance, dashed line: relative
904 abundance of NVP resistant strains. The respective half-life of NVP in the distinct
905 representative mothers was 1.3, 1.7, 2 and 2.6 days for panels B-E.

906 Figure 7: HIV-1 transmission risk in the case of extended newborn NVP dosing.
907 A: Predicted transmission risk after 6 weeks extended NVP treatment (solid line) and
908 confidence range (light-grey area) together with clinical data from the SWEN-study (3,
909 38) (open circles). B: Predicted transmission risk after 6 month extended NVP treatment
910 (solid line) and confidence range (light-grey area) together with clinical data from the
911 HPTN 046-study (10) (open squares). The intrauterine transmission risk was assumed
912 to be 5% (12). C: Predicted transmission risk after 2 years, in the case of no prophylaxis,
913 a single dose maternal and newborn NVP dose, 6-, 14-, 21 weeks, 6 month, 52-, 78- or
914 104 weeks of extended newborn NVP in addition to a single intrapartum maternal NVP
915 dose.

916 **Tables**

917 Table 1: Virus dynamics parameters. All units in [1/day], except the point mutation
 918 probability μ in [1/rev. transcr./base], the infection rate constant $\beta(wt, \phi)$ in [1/virions/day]
 919 and the T-cell production λ [cells/day/kg body weight].

Parameter	Value	Reference
k_T	0.35	(62)
δ_{T2}	1	(31)
N	1000	(45)
μ	$2.16 \cdot 10^5$	(30)
δ_T, δ_{T1}	0.02	(45)
$\beta(wt, \phi)$	$8 \cdot 10^{-12}$	(45)
δ_{PIC}	0.35	(55)
λ (newborn)	Eq. S2 [§]	
CL _V (newborn)	Eq. S4 [§]	
λ (mother)	$2.86 \cdot 10^7$ *	(57)
CL _V (mother)	23	(31)

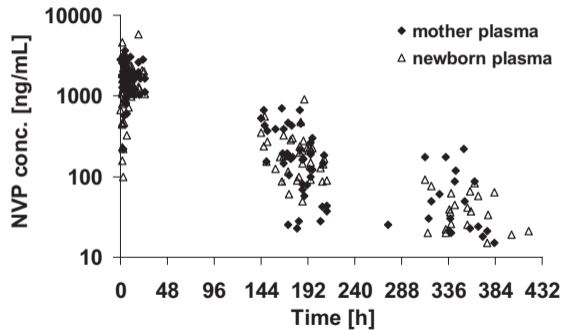
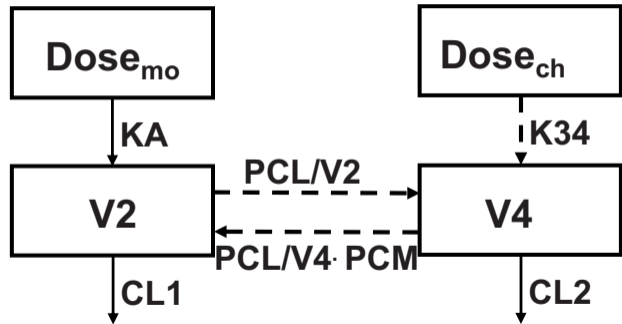
920 *The maternal zero-order T-cell production of $2 \cdot 10^9$ (56) was divided by the weight (70 kg)
 921 of the patients in (56), to yield the parameter stated in the table. [§] see supplementary
 922 text S1.

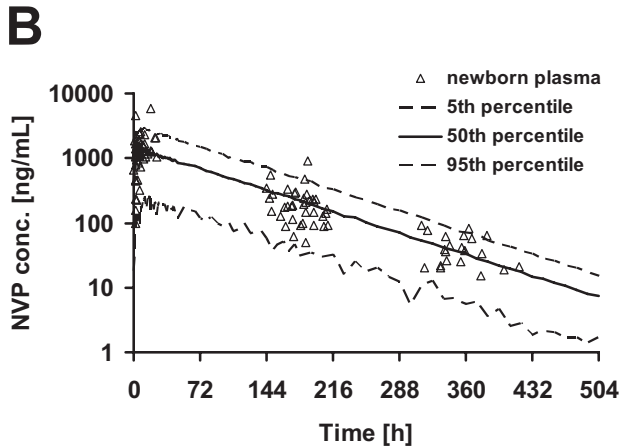
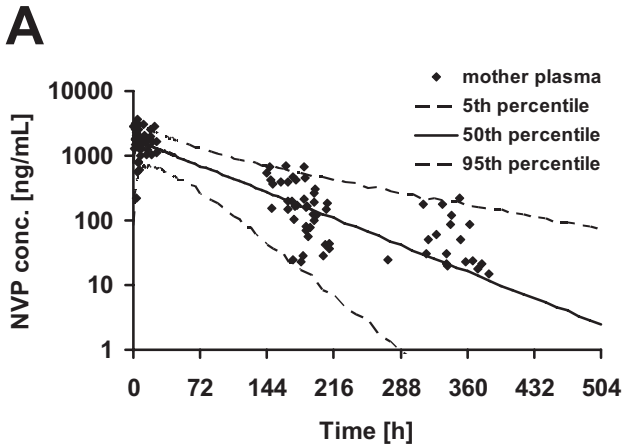
923 Table 2: Population PK estimates of NVP of the final combined PK model for mothers
 924 and newborns including results of the bootstrap.

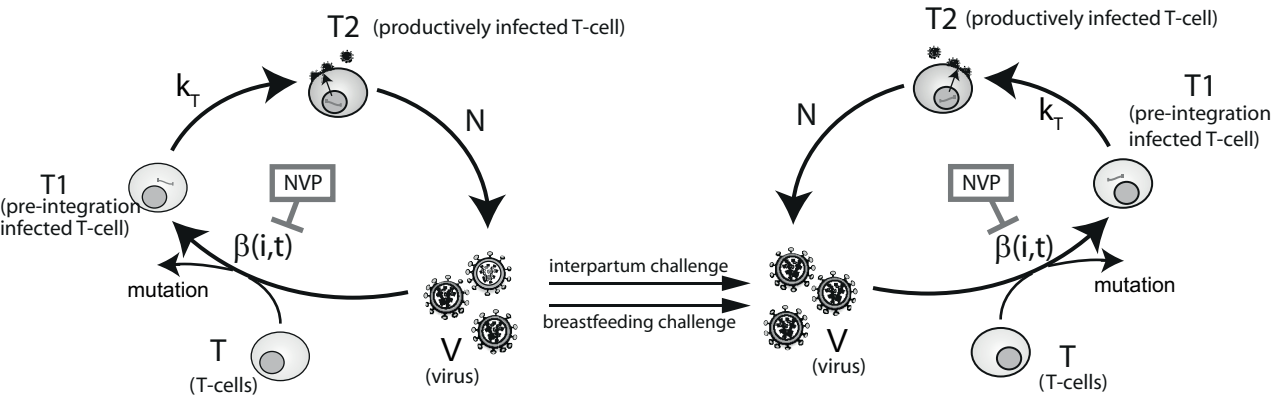
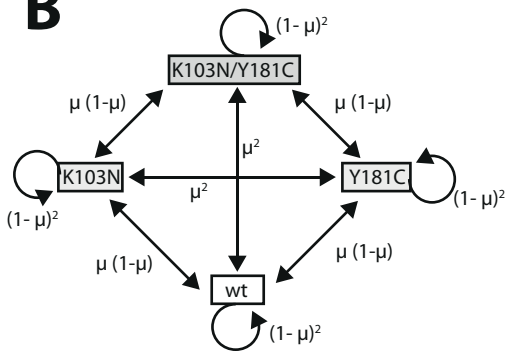
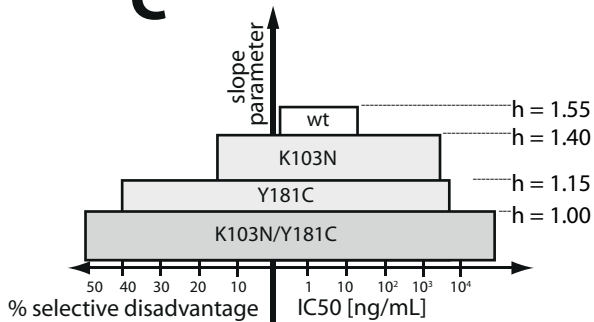
Model parameters	Units	Population estimates	RSE^a, %	Bootstrap^b median	95% Confidence interval (2.5th and 97.5th percentile)
<i>FIXED EFFECTS</i>					
KA	[h ⁻¹]	1.34 fixed	-	-	-
V2/F	[L]	90.9	5.85	89.6	75.4 – 101.4
CL1/F	[L·h ⁻¹]	1.22	6.33	1.20	1.01 – 1.38
V4/F	[L]	20.0	18.6	20.0	9.92 – 37.2
CL2/F	[L·h ⁻¹]	0.21	16.1	0.21	0.11 – 0.38
K34	[h ⁻¹]	1.34 fixed	-	-	-
PCL/F	[L·h ⁻¹]	111.0	20.5	99.7	5.90 – 463.8
PCM		1.38	7.68	1.36	1.10 – 1.61
<i>RANDOM EFFECTS</i>					
<i>Interindividual Variability</i>					
ωKA	[% CV]	159.7	30.3	150.5	51.9 – 209.8
ωCL1/F	[% CV]	32.9	25.7	31.8	23.1 – 40.5
ωV2/F	[% CV]	34.1	33.1	33.1	20.7 – 43.4
<i>Residual Variability</i>					
σ proportional (mothers)	[% CV]	27.2	10.6	26.8	19.4 – 32.1
σ proportional (newborns)	[% CV]	49.1	11.0	48.1	38.9 – 59.0

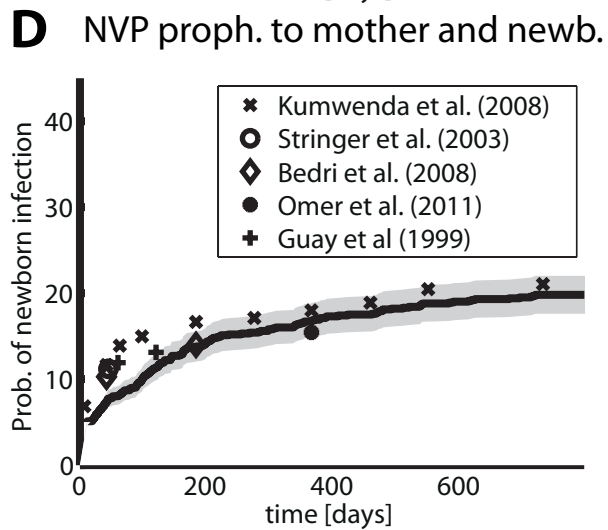
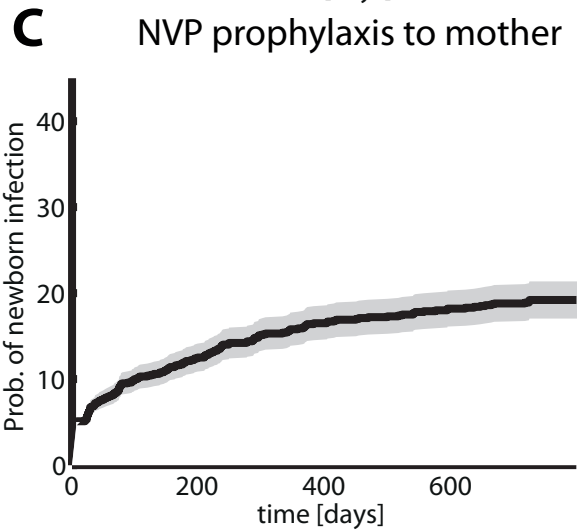
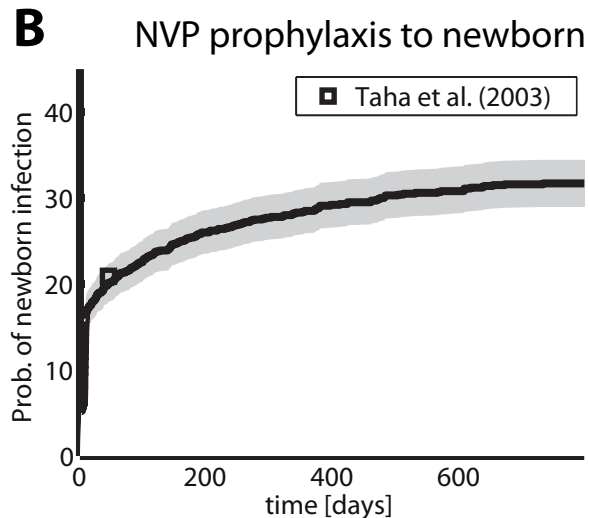
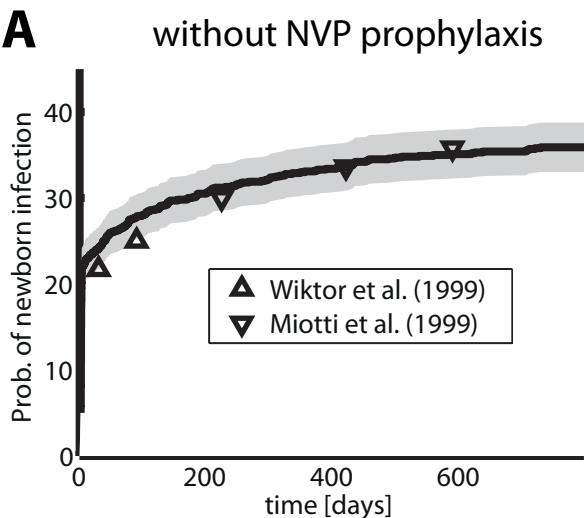
925 ^a Relative standard error (standard error divided by population estimate ·100;
 926 for the random effects parameters RSE is related to the corresponding variance scale).

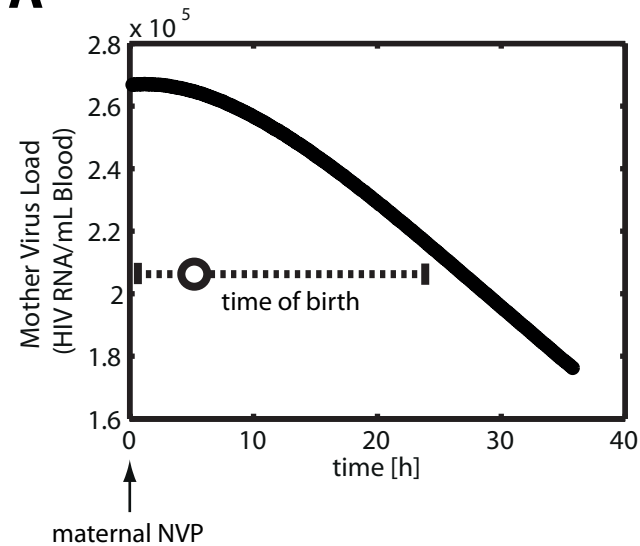
927 ^b n=1668

A**B**



Mother**Newborn****A****B****C**



A**B**